



**THE MICRONUCLEUS ASSESSMENT OF BUCCAL MUCOSA: A NONINVASIVE
METHOD IN SCREENING OF SMOKERS POTENTIALLY EXPOSED TO ORAL
CANCER**

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Key Words: Micronucleus; smoking; cancer; oral; Buccal mucosa

ABSTRACT

Background: Micronucleus is a chromatin structure in cytoplasm which can be an indicator of chromosome damage and carcinogenesis. The present study assessed micronuclei in exfoliated oral mucosal cells of heavy smokers.

Methods: This historical cohort study was performed on 48 individuals (24 smokers and 24 non-smokers) who were selected according to gender and age. The age of non-smokers equaled 34.42 ± 16.38 and the age of smokers (case group) equaled 36.42 ± 16.17 . Buccal mucosa smear was obtained with a moist wooden spatula and after placing the smear on glass slides and Papanicolaou staining, 1000 cells of each sample were inspected under light microscope and cells with micronucleus were counted. Data were analyzed by T-test and level of significance was set as lower than 0.05.

Results: The percentage of micronucleus was 21.21 ± 2.18 and 3.5 ± 1.17 respectively and this difference was determined to be statistically significant based on T-test and with 99% confidence level. ($p < 0.01$)

Conclusion: The average frequency of micronucleus was higher in smokers compared to non-smokers in this study, therefore this noninvasive assessment may help in early diagnosis of oral cancer.

Key words: Micronucleus; smoking; cancer; oral; Buccal mucosa

INTRODUCTION

Oral cancer is one of the ten most common human cancers with 575000 new cases and 320000 mortalities per year worldwide. [1] The prevalence of oral cancer in the world is often correlated with the pattern of tobacco products consumption and a dose-response relationship exists between the level of consumption of tobacco products and prevalence of oral cancer [2]. Oral cancer has a complex karyotype which is accompanied by many types of chromosomal deletion, translocation and chromosome abnormalities. Often, an error occurs in chromosome separation so that chromosome or part of it is left behind from the nucleus and remains in the cytoplasm in anaphase which is called micronucleus. [3] Micronucleus assay in buccal mucosal cells has been used for evaluation of chromosome damages and can help in early diagnosis of malignancies with simple biopsy and minimum invasion without causing stress to individuals. The advantage of this method is its simplicity because micronucleus assay is quick and does not require any specific specialty. Kamboj et al. showed that evaluation of epithelial cells of buccal mucosa is a reliable marker in early diagnosis of premalignant and malignant lesions, [

4] in a way that assessment of micronuclei in exfoliated epithelial cells of mucosa has been proposed as a method for specification of cancer risk. [3,5,6] in another study in 2011, number of micronuclei in oral mucosal cells of malignant and potentially malignant groups was significantly higher than that of the control group and it has been proposed to use this method for prognosis screening and also as an instructive method in public centers. [7] This genomic damage can occur due to exposure to genotoxic substances, medical interventions (radiation and chemical substances), micronutrients deficiency (folic acid), life style (alcohol, cigarettes, narcotics and stress), air pollution, chronic contact with arsenic and chromium and genetic factors such as defect in DNA metabolism or repair. [8] Because of the importance of micronucleus assay, Human Micronucleus Project (HUMN) was held in 1997 to standardize the assessment of this index in peripheral blood lymphocytes and the results of this project have been published [9, 10, 11]. Based on our knowledge from available resources, till now only few studies assessed the diagnostic value of the mentioned method in smokers. [12] Therefore, considering th

the importance of early diagnosis of cancers, the present study was performed to evaluate the relation between cigarette smoking and number of micronuclei and to compare the results with that of non-smokers.

MATERIALS AND METHODS

This study was performed on 48 individuals including 24 subjects in the control group (non-smokers) and 24 subjects in the case group (smokers) with a history of smoking for more than 5 years and more than 20 cigarettes per day which are considered as heavy smokers. [13] Subjects in both groups were males residing in Tehran and were similar regarding socio-economic status and time of assessment and were homogenized regarding age. Subjects with recent viral disease, medicine consumption, alcohol consumption, addiction to narcotics, high risk occupations such as paint industry and radiotherapy were excluded i.e. selected smoking subjects only smoked cigarettes and non-smokers were completely normal.

After receiving an informed consent, subjects were asked to rinse their mouth with water. Buccal mucosal cells were scrapped by a moist spatula under medium pressure and were placed on clean glass slides and were fixed with path fix spray. Standard Papanicolaou staining was used for micronucleus assay. 1000 Cells were inspected under light microscope (Nikon, Japan) with $\times 400$ and $\times 1000$ magnification. The criteria by Tolbe

rt et al. were adapted for micronucleus count, [14, 15] which consists of the followings:

- 1- Round and smooth periphery which indicates a membrane.
- 2- Less than 1/3 of the diameter of the nucleus but big enough for distinction of shape and color.
- 3- Color intensity similar to the nucleus.
- 4- Consistency and texture similar to the nucleus.
- 5- Focal length similar to the nucleus.
- 6- No connection or overlapping with the nucleus.

Cells with distinctive margins and nuclei were counted. Micronuclei were not counted in areas with cell overlap. Also, dead and degenerated cells and nuclear bubbles were excluded from cell count. Finally, data were analyzed by T-test and level of significance was set as lower than 0.05.

RESULTS

The age of non-smokers equaled 34.42 ± 16.38 and the age of smokers (case group) equaled 36.42 ± 16.17 . Chi-square test showed that the two groups have no significant age difference. T-test was used for statistical analysis and $p < 0.01$ was considered as statistically significant. Mean, standard deviation and mean difference of data were calculated. The frequency of micronucleus are presented in table 1 based on case and control groups and show that the minimum percentage of micronucleus is in non-smokers (2.4) and this value equaled 9.9 in smokers with statistical significant difference between the two groups. ($p < 0.01$). Figure 1 shows cell w

it-hmicronucleus.

The average frequency of micronucleus in study groups is presented in table 2.

The percentages of micronucleus per 1000 counted cells divided by case and control groups are presented in diagram 1.

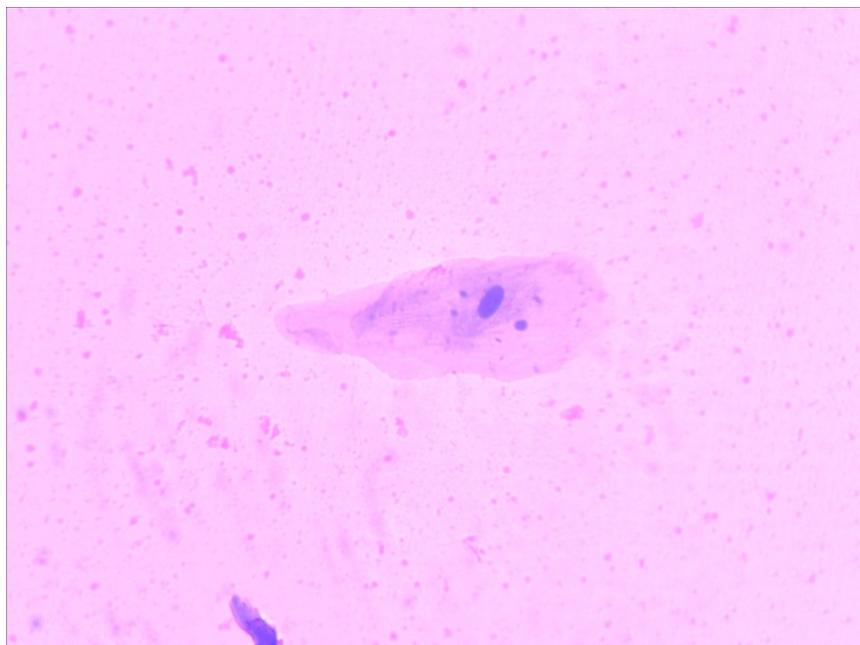


Figure 1 – Micronucleus in Papanicolaou staining (×1000 magnification)

Table 1 – Percentage of micronucleus based on case and control groups

Micronucleus / Cigarette smoking	Amount	Minimum-Maximum	Variation coefficient
Non-smoker (control) (N=24)	3.5±1.17	2.4 – 6.1	33.43
Smoker (case) (N=24)	12.12±2.18	9.9 – 18.5	17.99

Table 2 – Average number of micronucleus in study groups

Group	Sample size	Average number of micronucleus per 1000 counted cells in each sample	T-test result P< .01
Smoker	24	121.25	Significant
Control	24	35	

DISCUSSION

Considering the heavy expenses of therapeutic and rehabilitating procedures of malignant lesions, the path of recent studies has been directed towards prevention and early diagnosis of these lesions. In this

regard, the present study assessed micronucleus frequency in buccal mucosa of heavy smokers and showed that the average percentage of micronucleus in smokers was significantly higher than that of non-smokers. This finding is in line with

previous studies which had shown that smoking elevates micronucleus frequency. [2, 3, 5, 12, 16, 18]. In other studies, smoking either had no effect on micronucleus frequency (20) or even decreased micronucleus in some cases. [21] These studies were performed on peripheral blood lymphocytes and it should be considered that the level of effective chemical substances in cigarettes may be lower in peripheral blood compared to other organs. In addition, smoking can damage lymphocytes in a way that they will be unable to survive or lose the ability to divide during stages of cell culture with cytokinesis-block micronucleus (CBMN) method. [11] In our study, smokers with a history of at least 5 years of smoking and at least 20 cigarettes per day which are considered as heavy smokers [1] were evaluated to better specify the effect of toxic substances of cigarette on production of micronucleus. In a comprehensive study by Bonassi et al. micronucleus frequency in peripheral blood lymphocytes of light or moderate smokers showed no significant increase. While in heavy smokers this increase was significant. [11] Bonassi et al. mentioned that in 33 researches among 37 published articles, no correlation was found between cigarette smoking and micronucleus increase. But in the majority of these studies heavy smokers have not

been considered which can justify the neutral effect of cigarette smoking in these studies. [11] But in our study, smokers had smoked at least 20 cigarettes per day for more than 10 years. Nevertheless, note that the level of effective chemical substances of cigarette in peripheral blood may be lower than other organs and this can justify the lack of increase in micronucleus frequency in light smokers when micronucleus assay is performed on peripheral blood cells [11]. JalayerNaderi et al. performed a study in 2012 entitled micronucleus assay of buccal mucosa cells in smokers of Iranian population with a history of smoking less and more than 10 years and compared the results with that of non-smokers and used Feulgen staining. They concluded that the percentage of micronucleus and also the average number of micronuclei in each cell in non-smokers are significantly lower than that of smokers. Although, the average number of micronuclei of oral mucosa cells was higher in smokers with more than 10 years of smoking compared to less than 10 years of smoking, but this difference was not significant. [12] Kamath et al. showed that individuals with a history of 5 to 10 years of smoking had more micronuclei than individuals who had smoked more than 10 years or less than 5 years. [2] But no such correlation was seen in our study. In the

present study, for example a 26-year-old individual who had smoked for 12 years had 185 micronucleus but a 73-year-old individual with a history of 40 years of smoking had 132 micronucleus. This difference can be due to the level of cell resistance against toxic elements in different individuals and also due to harmful factors other than cigarettes which of course this issue needs more evaluation. In addition, it is worth mentioning that the present study was performed on heavy smokers, while in other studies either heavy smoking criteria were not mentioned or subjects were not heavy smokers. Nevertheless, Wu et al. showed that a significant correlation exists between number of cigarettes per day and number of micronuclei and heavy smokers have more micronuclei in oral mucosa [22]. The applied staining methods for micronucleus specification in exfoliated oral mucosal cells have been different in various studies and include Feulgen-Fast green, fluorescent staining such as Diamidino-2-phenylindole DAPI, Hoechst, Acridine orange, Propidium iodide, May-Grunwald Giemsa and Papanicolaou. [1] It seems that Feulgen-Fast green staining is of higher interest among researchers due to its specificity for DNA and clear staining of cytoplasm which eases micronucleus count. [1] Nevertheless, Papanicolaou staining has

been used in many recent studies [2, 3, 23] including the present study with acceptable results. This method neither needs complex procedures such as cell culture or preparation of metaphase nor requires DNA-specific staining [8]. In other studies, mostly cigarettes and other forms of tobacco have been compared or study participants were using multiple substances while in the present study, only the effect of cigarette smoking on the average number of micro nucleated cells was assessed. Because based on the results of some researches when participants use multiple substances, for example cigarette and chewing tobacco, interaction between these substances causes nucleus degeneration and an appearance similar to micronucleus structure which affects the study results. [12] In the present study, confounding factors especially alcohol consumption were eliminated and the synergic effect of these two substances on the number of micronuclei was omitted and in this way, significant increase in number of micronuclei in smokers was more decisively correlated to cigarette smoking. It is clear that the synergic effect of cigarette smoking and alcohol consumption reaches to more than 5.5-fold [24]. Bansal et al. assessed micronuclei in tobacco consumers of Indian Punjabi population in 2011 with Papanicolaou staining method

and concluded that micronucleus in smokeless tobacco consumers are significantly more than that of smokers and the control group. Also, the number of micronuclei was higher in smokers compared to the control group. [3] Studies performed by Motgi et al. [23], Patel et al. [25] and Palaskar et al. [26] also revealed similar results and indicated the higher carcinogenicity of smokeless tobacco compared to cigarette. In another study, Caplash et al. assessed micronucleus in exfoliated oral mucosa cells of chewing tobacco users, smokers and the control group. They concluded that micronucleus frequency in tobacco consumers was significantly higher than the control group. Two interfering factors of age and exposure duration showed a significant correlation, while the third interfering factor (alcohol) showed no significant correlation. [19] On the other hand, in some studies controversies are seen among the results regarding the gender of patients in a way that some studies reported higher frequency of micronucleus in males and others reported it higher in females. [27-29] This controversy also exists regarding the age of individuals. [17, 28, 30, 31] In the present study, all subjects were males and were homogenized regarding their age to omit the effect of age and gender on the results. Although Sarto et al. reported that

micronucleus frequency due to chromosome breakage is doubled in smokers compared to non-smokers [16] but in our study this amount was approximately 3.5-fold. This difference and other mentioned differences can be due to different staining methods, sample sizes and assessment of different smokeless and smoking substances. Note that the chemical composition of cigarettes of different manufacturers and that of smokeless substances are highly different. Therefore, more definite results require standardizing of applied methods. All mentioned studies and the present study show that unusual oral habits increase micronucleus (4) In other words, micronucleus assay can be applied as a quick and simple index and a valid biomarker for early diagnosis of premalignant and malignant oral lesions. Nevertheless, further researches with more precise techniques are required in this regard.

CONCLUSION

This study showed that the frequency of micronucleus in buccal mucosa of heavy smokers is significantly higher than that of non-smokers with 3.5-fold increase. ($p < 0.01$) therefore, this noninvasive assessment may help in early diagnosis of oral cancer. Nevertheless, to reach justifiable and reliable results, larger groups should be assessed for longer time periods. Usually, a limited number

of patients are willing to reveal duration and frequency of cigarette smoking and other habits. Also, buccal cells should be reassessed after habit weans to see whether or not micronuclei have decreased in number. In addition, other genetically harmful factors in the living environment of each individual should also be considered.

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CONFLICT OF INTEREST:

The authors have no conflict of interest in this study.

REFERENCES

- 1-Kashyap B, Reddy PS. Micronuclei assay of exfoliated oral buccal cells: Means to assess the nuclear abnormalities in different diseases. *J Can Res Ther* 2012;8(2):184-91
- 2-Kamath VV, Anigol P, Setlur K. Micronuclei as prognostic indicators in oral cytological smears: A comparison between smokers and non-smokers. *Clin Cancer Investig J* 2014;3(1):49-54
- 3-Bansal H, Sandhu V S, Bhandari R, Sharma D. Evaluation of micronuclei in tobacco users: A study in Punjabi population. *ContempClin Dent* 2012; 3(2): 184–7
- 4-Kamboj M, Mahajan S. Micronucleus--an upcoming marker of genotoxic damage. *Clin Oral Investig* 2007;11(2):121-6.
- 5-Stich HF, Stich W, Parida BB. Elevated frequency of micronucleated cells in the buccal mucosa of individuals at high risk for oral cancer: betel quid chewers. *Cancer Lett.* 1982 ;17(2):125-34.
- 6-Cairns J. Mutation selection and the natural history of cancer. *Nature* 1975;255:197–200
- 7- Devi P, Thimmarasa VB, Mehrota V, Arora P. Micronucleous assay for evaluation of genotoxicity in potentially malignant and malignant disorders. *JIAOMR* 2011;23:97-100
- 8-Huen K, Gunn L, Duramad P, Jeng M, Scalf R, Holland N. Application of a geographic information system to explore associations between air pollution and micronucleus frequencies in African American children and adults. *Environ Mol Mutagen* 2006;47(4):236-46.
- 9-Bonassi S, Fench M, Lando C, Lin YP, Ceppi M, Chang WP, et al. Human Micronucleus project: International database comparison for results with the cytokinesis block micronucleus assay in human lymphocytes: I Effect of laboratory protocols, scoring criteria and host factors on the frequency of micronuclei. *Environ Mol Mutagen* 2001;37:31-45.
- 10- Bonassi S, Nerri M, Puntoni R. Validation of biomarkers as early predictors of disease. *Mut Res* 2001;480:349-58.
- 11- Bonassi S, Neri M, Lando C, Ceppi M, Lin YP, Chang WP. Effect of

- smoking habit on the frequency of micronuclei in human lymphocytes: results from the Human MicroNucleus project. *Mutat Res* 2003;543(2):155-66.
- 12- Naderi NJ, Farhadi S, Sarshar S. Micronucleus assay of buccalmucosa cells in smokers with the history of smoking less and more than 10year s. *Indian J PatholMicrobiol* 2012;55(4):433-8.
- 13-Baumert J, Ladwig KH, Ruf E, Meisinger C, Döring A, Wichmann HE, etal. Determinants of heavy cigarette smoking: are there differences in men and women? Results from the population-based MONICA/KORA Augsburg surveys. *Nicotine Tob Res.* 2010 Dec;12(12):1220-7.
- 14- Tolbert PE, Shy CM, Allen JW. Micronuclei and other nuclear anomalies in buccal smears: Methods development. *Mutat Res* 1992;271(1):69-77.
- 15-Tolbert PE, Shy CM, Allen JW. Micronuclei and other nuclear anomalies in buccal smears: A field test in snuff users. *Am J Epidemiol* 1991;134(8):840-50.
- 16- Sarto F, Finotto S, Giacomelli L, Mazzotti D, Tomanin R, Levis AG. The micronucleus assay in exfoliated cells of the human buccalmucosa. *Mutagenesis* 1987;2(1):11-7.
- 17-Ozkul Y, Donmez H, Erenmemisoglu A, Demirtas H, Imamoglu N. Induction of micronuclei by smokeless tobacco on buccal mucosa cells of habitual users. *Mutagenesis* 1997;12(4):285-7.
- 18-Haveric A, Haveric S, Ibrulj S. Micronuclei frequencies in peripheral blood and buccal exfoliated cells of young smokers and non-smokers. *ToxicolMech Methods* 2010 ;20(5):260-6.
- 19- Caplash S, Meenakshi. MICRONUCLEUS INVESTIGATION IN EXFOLIATED BUCCAL CELLS AMONG TOBACCO CHEWERS/ SMOKERS AND CONTROLS. *IJBPAS* 2013; 2(1): 72-79.
- 20- Yager JW, The effect of background variables on human peripheral lymphocyte micronuclei. *IARC Sci Publ*1990; 104:147-50.
- 21-Barale R, Chelotti L, Davini T, Del Ry S, Andreassi MG, Ballardini M, etal. Sister chromatid exchanges and micronucleus frequency in human lymphocytes of 1650 subjects in an Italian population. II. Contribution of sex, age, and lifestyle. *Environ Mol Mutagen* 1998;31(3):228-42.
- 22-Wu PA, Loh CH, Hsieh LL, Liu TY, Chen CJ, LiouSH. Clastogenic effect for cigarette smoking but not arecaquid chewing as measured by micronuclei in exfoliated buccal mucosal cells. *Mutat Res* 2004;562(1-2):27-38.
- 23- Motgi AA, Chavan MS, Diwan NN, Chowdhery A, Channe PP, Shete MV.

Assessment of cytogenic damage in the form of micronuclei in oral epithelial cells in patients using smokeless and smoked form of tobacco and non-tobacco users and its relevance for oral cancer. *J Cancer Res Ther* 2014;10(1):165-70.

24- Stich HF, Rosin MP. Quantitating the synergistic effect of smoking and alcohol consumption with the micronucleus test on human buccal mucosa cells. *Int J Cancer* 1983;31(3):305-8

25- Patel BP, Trivedi PJ, Brahmabhatt MM, Shukla SN, Shah PM, Bakshi SR. Micronuclei and chromosomal aberrations in healthy tobacco chewers and controls: A study from Gujarat, India. *Arch Oncol* 2009;17(1-2):7-10.

26- Palaskar S, Jindal C. Evaluation of micronuclei using papanicolaou and May grunwaldgiemsa stain in individuals with different tobacco habits- A comparative study. *J ClinDiagn Res* 2010;4:3607-3613.

27- Gonsebatt ME, Vega L, Salazar AM, Montero R, Guzmán P, Blas J, et al. Cytogenetic effects in human exposure to arsenic. *Mutat Res* 1997;386(3):219-28.

28- Piyathilake CJ, Macaluso M, Hine RJ, Vinter DW, Richards EW, Krumdieck CL. Cigarette smoking, intracellular vitamin deficiency, and occurrence of micronuclei in epithelial cells of the buccal mucosa. *Cancer Epidemiol Biomarkers Prev* 1995;4(7):751-8.

29-Benites CI, Amado LL, Vianna RA, Martino-Roth Mda G. Micronucleus test on gas station attendants. *GenetMol Res* 2006;5(1):45-54.

30- Konopacka M. Effect of smoking and aging on micronucleus frequencies in human exfoliated buccal cells. *Neoplasma* 2003;50(5):380-2

31- Bohrer PL, Filho MS, Paiva RL, da Silva IL, Rados PV. Assessment of micronucleus frequency in normal oral mucosa of patients exposed to carcinogens. *ActaCytol* 2005; 49(3):265-72.